

B93 CCGCCA^GCCCCA (SEQ ID NO: 20)
B98 AGCCC^CAGGGAA (SEQ ID NO: 21)
B227 TGAGCT^{CTGTC}AACGCC (SEQ ID NO: 22)
C82 TGGAGT^{TGGA..37bp.GAGT}GGATTG (SEQ ID NO: 23)
C209 AGCACC^{TCTCCCTGAAGTT}GAGCTC (SEQ ID NO: 24)
C187 ATATCA^{GTACACAGTCCAAGA}AGCACC (SEQ ID NO: 25)
U26 CGGAGA^{CC}CTGCC (SEQ ID NO: 26)
U199 ACGTCC^{AAG}AAGCAC (SEQ ID NO: 27)
U208 AAGCAG^CTTTCTC (SEQ ID NO: 28)
U268 GCGAGA^{GTTATTA}CTAGGG (SEQ ID NO: 29)

A255 TGTGCGAGAGTTATTACGAGAGTTATTA^{CTAGGG} (SEQ ID NO: 30)
A113 GGCTGGAGTGGATTGGG.62bp.TATCAGTGGATTGGG.62bp.TATC^{AGTAGA}
(SEQ ID NO: 31)
U43 ACCTGCGGTGTTTATGGTGTTTAT^{GGTGGG} (SEQ ID NO: 32)
U318 GGACGTCTGGGGCCAACGTCTGGGGCCA^{AGGGAC} (SEQ ID NO: 33)
D27 GGAGAC^{CCTCA}CCTGCG (SEQ ID NO: 34)
D31 ACCCTC^ACCTGCG (SEQ ID NO: 35)
D219 CCTGAA^GTTGAGC (SEQ ID NO: 36)
D150 CACCAA^CTACAAC (SEQ ID NO: 37)
D109 AAGGGG^CTGGAGT (SEQ ID NO: 38)
E28 CCCTCA^{CCTGC}GGTGTT (SEQ ID NO: 39)
E81 CTGGAG^{TTGGA..37bp..TGGAG}TGGATT (SEQ ID NO: 40)
E88 TGGATC^{CGCC}AGCCCC (SEQ ID NO: 41)
E92 CGCCA^GCCCCCA (SEQ ID NO: 42)
E136 AATCAT^{AGTGGAAGCACCAACTA}CAACCC (SEQ ID NO: 43)

A2
cont.

- F66 CTTAC^{TGGTTACTACT}GGAGTT (SEQ ID NO: 44)
- F183 TATCAT^{ATCAGTA}ACACGT (SEQ ID NO: 45)
- F215 TCTCCC^{TGAA.18bp.CGCC}GCGGAC (SEQ ID NO: 46)
- F267 TGCGAG^{AG}TTATTA (SEQ ID NO: 47)
- D55 TATGGTGG.41bp.AGGGGTGG.41bp.AGGG^{AAGG} (SEQ ID NO: 48)
- D123 GATTGGGGAAATCAATCATAGTGGAAGCGGAAATCAATCATAGGGAAGC^{ACCAAC}
(SEQ ID NO: 49)
- F85 AGTTGGAT.10bp.CCCAGGAT.10bp.CCCA^{GGGA} (SEQ ID NO: 50)
- D3 GGTCGC^{AGGACTGT}_{GACCC}TGAAGC (SEQ ID NO: 51)
- D56 ATGGTGGG.50bp.CAGGGGGTGGG.50bp.CAGGG^{AAGGGG} (SEQ ID NO: 52)
- D71 GTGGTT^A_{GGG}CTACTG (SEQ ID NO: 53)
- D75 TTACTA^C_{GG}TGGAGTT (SEQ ID NO: 54)
- D126 TGGGA^{AATCAATCAT}_{GGG}AGTGGA (SEQ ID NO: 55)
- D223 AAGTTG^{AG}_{GACCCGGC}CTCTGTG (SEQ ID NO: 56)
- D232 TCTGTGAACGCCGC^{GGACAC}GCCCCGTCCTGTGAACGCCGC (SEQ ID NO: 57)
- D235 GTAAAC^{GGAGG}_{GCCGCG} (SEQ ID NO: 58)
- D252 GGCTGT^{GTATTACTGT}_{TCC}GCGAGA (SEQ ID NO: 59)
- D268 GCGAGA^{GT}_{AGG}TATTATT (SEQ ID NO: 60)
- D275 TTATTA^C_{GG}TAGGGC (SEQ ID NO: 61)
- D332 AAGGGA^C_{AG}CAC (SEQ ID NO: 62)
- E3 GGGCGC^{AGGA.51bp.CTTC}_{GT}AGTGGT (SEQ ID NO: 63)

A2
cont.

- E51 TGT^TTTA^{TGGT.15bp.TACT}ACTGGAG (SEQ ID NO: 64)
AGACC
- E80 ACTGGA^GTTGGAT (SEQ ID NO: 65)
CCC
- E263 ACTGTG^{CGAGAGTTATTACT}AGGGCG (SEQ ID NO: 66)
GGTG
- F89 GGATCC^{GCCAGCCCCAGGG}AAGGGG (SEQ ID NO: 67)
AGG
- F168 CCTCAA^{AGAGTCGAGT}CACCAT (SEQ ID NO: 68)
GGG
- F195 AGACAC^{GTCCAAGAAG}CACCTC (SEQ ID NO: 69)
AGGGC
- F199 ACGTCC^{AAGAAG}ACCCTGA (SEQ ID NO: 70)
CT
- F242 CCGCGG^{ACACGGCTGTGTATTACTGT}GCGAGA (SEQ ID NO: 71)
GGA
- F260 ATTACT^{GTG}CGAGAG (SEQ ID NO: 72)
CGTGA
- F264 CTGTGC^{GAGAG.46bp.CGTC}TGGGGC (SEQ ID NO: 73)
ACA
- B123 GATTGG^GAAAATC (SEQ ID NO: 74)
- C109 AAGGGT^CTGGAGT (SEQ ID NO: 75)
- A16 TTGAAGCCTTCGGACT^{GAAGCCTTCGGAGA}CCCTGT (SEQ ID NO: 76)
- U180 AGTCACCATATCAA^{ACCATATCAG}TAGACA (SEQ ID NO: 77)
- D45 CTGCGCG^{GTTTATGGTGGGT}CCTTCA (SEQ ID NO: 78)
- D164 CGTCCCC^{CAAG}AGTCGA (SEQ ID NO: 79)
- D216 CTCCCTT^{AAG.22bp.CGGA}CACGGC (SEQ ID NO: 80)
- E11 GACTGT^TAAAGCC (SEQ ID NO: 81)
- E54 TTATGGA^{GGG.25bp.GTTG}GATCCG (SEQ ID NO: 82)
- F188 TATCAGG^{AGACACGTCCAGAA}GCACCT (SEQ ID NO: 83)
- F220 CTGAAGC^{TGAGCTCTGTG}AACGCC (SEQ ID NO: 84).--

A2
cont.

3. On page 8, please replace the paragraph from lines 3-6 with the following:

A3
--Figure 7 provides a comparison of sequences isolated from VH genes of Ramos cells which have lost anti-idiotypic (anti-Id1) binding specificity. Nucleotide substitutions which differ from the starting population consensus are shown in bold. Predicted amino acid changes are indicated, also in bold type. The nucleotide sequence of the VH Ramos gene depicted is SEQ ID NO: 15.--

4. On page 8, please replace the paragraph from lines 13-15 with the following:

A4
--Figure 11 shows a V_H sequence (SEQ ID NO: 15) derived from streptavidin-binding Ramos cells. Figure 11 CONT'D shows the V_L sequence of SEQ ID NO: 85. Nucleotide changes observed in comparison with the V_H sequence of the starting population, and predicted amino acid changes, are shown in bold.--

5. On page 8, please replace the paragraph from line 24 with the following:

A5
--Figure 16 shows V_H (SEQ ID NO: 15) and V_L (SEQ ID NO: 85) sequences of round 6 selected IgM.--

6. On page 9, please replace the paragraph from lines 9-10 with the following:

A6
--Figure 21 provides an analysis of Ig sequences of unsorted DT40 populations after one month of clonal expansion. The sequence of Figure 21 d is SEQ ID NO: 86.--

7. On page 9, please replace the paragraph from lines 13-14 with the following:

A7
--Figure 23 provides an analysis of naturally-occurring constitutively hypermutating BL cell lines. The sequence of Figure 23 C is SEQ ID NO: 87.--

8. Please replace the paragraph on page 23, lines 20-28 extending to page 24, lines 1-14 with the following replacement paragraph:

A8
--In order to screen for a cell that undergoes hypermutation *in vitro*, the extent of diversity that accumulates in several human Burkitt lymphomas during clonal expansion is assessed. The Burkitt lines BL2, BL41 and BL70 are kindly provided by G. Lenoir (IARC, Lyon, France) and Ramos (Klein *et al.*, 1975, *Intervirology* 5: 319-334) is

A8
CONT.

provided by D. Fearon (Cambridge, UK). Their rearranged V_H genes are PCR amplified from genomic DNA using multiple V_H family primers together with a J_H consensus oligonucleotide. Amplification of rearranged V_H segments is accomplished using Pfu polymerase together with one of 14 primers designed for each of the major human V_H families (Tomlinson, 1997, V Base database of human antibody genes. Medical Research Council, Centre for Protein Engineering, UK. <http://www.mrc-cpe.cam.ac.uk/>) and a consensus J_H back primer which anneals to all six human J_H segments (JOL48, 5'-GCGGTACCTGAGGAGACGGTGACC-3' (SEQ ID NO: 1), gift of C. Jolly).

Amplification of the Ramos V_H from genomic DNA is performed with oligonucleotides RVHFOR (5'-CCCCAAGCTTCCCAGGTGCAGCTACAGCAG (SEQ ID NO: 2),) and JOL48. Amplification of the expressed V_H-C_μ cDNA is performed using RVHFOR and C_μ 2BACK (5'-CCCCGGTACCAGATGAGCTTGGACTTGCGG (SEQ ID NO: 3)). The genomic C_μ C1/2 region is amplified using C_μ 2BACK with C_μ 1FOR (5'-CCCCAAGCTTCGGGAGTGCATCCGCCCAACCCTT (SEQ ID NO: 4)); the functional C_μ allele of Ramos contains a C at nucleotide 8 of C_μ 2 as opposed to T on the non-functional allele. Rearranged V_λ's are amplified using 5'-CCCCAAGCTTCCCAGTCTGCCCTGACTCAG (SEQ ID NO: 5) and 5'-CCCCTCTAGACCACCTAGGACGGTC-AGCTT (SEQ ID NO: 6). PCR products are purified using QIAquick (Qiagen) spin columns and sequenced using an ABI377 sequencer following cloning into M13. Mutations are computed using the GAP4 alignment program (Bonfield *et al.*, 1995, *NAR* 23: 4992-99).--

9. On page 39, please replace the paragraph on lines 12-19 with the following replacement paragraph:

A9

--For the *tk-neo* insert in *tk-neo::C_κ* mice, the amplified region extends from residues 607 to 1417 [as numbered in plasmid pMCNeo (GenBank accession U43611)], and the nucleotide sequence determined from position 629 to 1329. The mutation frequency of endogenous VJ_κ rearrangements in *tk-neo::C_κ* mice is determined using a strategy similar to that described in Meyer *et al.*, 1996. Endogenous VJ_κ5 rearrangements are amplified using a V_κ FR3 consensus forward primer